

Biological Research Registration Form

The University of Oregon requires Institutional Biosafety Committee (IBC) oversight, review and approval of **research involving recombinant or synthetic nucleic acids (rsNA), transgenic organisms, Risk Group 2 organisms (RG 3 not permitted), biological toxins, select agents, and non-human primate materials. Recombinant research exempt from *NIH Guidelines*, research involving human and animal biospecimens or human and animal cell culture must still be registered with the UO IBC.** Please complete this registration form and submit to the UO Biosafety Officer for initial review. Typically, separate forms do not need to be completed for individual grants and single IBC registration may cover several aspects of research. Please remember to sign the form on the last page.

## SECTION I Date Approved:

|  |  |  |
| --- | --- | --- |
| **Principal Investigator (PI):** | | **Registration #** (issued by EHS)**:**  **2025-** |
| **College: Arts and Sciences** | **Department:** | |
| **Phone Number:** | **Email:** | |
| **Project Title:** | | |
| **Personnel working on project:** | | |
| **Lab Building(s) and Room Number(s):** | | |
| **Please list all applicable UO Sponsored Projects EPCS Record #s or Banner Grant #s:** | | |
| **Institutional Animal Care & Use Committee (IACUC) current approval number(s),**  **if applicable:** | | |
| **Institutional Review Board Protocol number(s),**  **if applicable:** | | |

***SECTION II:*** Please complete applicable questions as they pertain to the work you are conducting.

# Recombinant or synthetic nucleic acids (rsNA) in *E.coli K-12*

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. List the bacterial host species: Click or tap here to enter text.
  2. Gene(s) encoded by inserted DNA: Click or tap here to enter text.
  3. Biological origin of inserted DNA: Click or tap here to enter text.
  4. Will inserted gene(s) be expressed in the host bacterium?  Yes  No

If yes, what are the gene product effects (toxicity, physiological activity, oncogenic potential, or

ability to alter cell cycle): Click or tap here to enter text.

* 1. List vectors to be used: Click or tap here to enter text.
  2. List antibiotic resistance genes contained on these vectors: Click or tap here to enter text.
  3. Do experiments involve large scale (>10 liters in one container) culture?  Yes  No
  4. Do experiments involve cloning toxin genes?  Yes  No

If yes, the Name of toxin: Click or tap here to enter text.

Do experiments involve transfer of antibiotic resistance gene in addition to those contained in the vectors?  Yes  No

If yes, the Name of antibiotic resistance gene.Click or tap here to enter text.

# Recombinant or synthetic nucleic acids (rsNA) in bacterial hosts other than E.coli K-12

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. List the bacterial host species: Click or tap here to enter text.
  2. Gene(s) encoded by inserted DNA: Click or tap here to enter text.
  3. Biological origin of inserted DNA: Click or tap here to enter text.
  4. Will inserted gene(s) be expressed?  Yes  No

If yes, what are the gene product effects (e.g., toxicity, physiological activity, oncogenic potential, ability to alter cell cycle): Click or tap here to enter text.

* 1. List Vectors to be used: Click or tap here to enter text.
  2. List antibiotic resistance genes contained on these vectors: Click or tap here to enter text.
  3. If the host is a human or animal pathogen, are any of the antibiotics listed above used to treat the disease in humans or animals:  Yes  No

If yes, are any of the antibiotics used to treat the disease in any patient population? Yes No

If yes, are any of the antibiotics used to treat the disease in other countries?  Yes No

If yes, list antibiotics that can be used to treat an infection with this microorganism, taking into consideration the inserted and naturally-occurring drug resistance genes: Click or tap here to enter text.

* 1. Do experiments involve large scale (> 10 L in one container) culture?  Yes No
  2. Do experiments involve cloning biological toxin gene?

If yes, name of toxin: Click or tap here to enter text. Do experiments involve transfer of an antibiotic resistance gene into the host in addition to those contained in the vectors?  Yes No

If yes, list the name of antibiotic resistance gene: Click or tap here to enter text.

# Recombinant or synthetic nucleic acids (rsNA) in lower eukaryotic hosts (e.g., parasites, fungi, algae)

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Name of host organism(s): Click or tap here to enter text.
  2. Gene(s) encoded by inserted DNA: Click or tap here to enter text.
  3. Biological origin of inserted DNA: Click or tap here to enter text.
  4. List Vectors to be used: Click or tap here to enter text.
  5. Will inserted gene(s) be expressed?  Yes  No

If yes, what are the gene product effects (e.g., toxicity, physiological activity, oncogenic potential, ability to alter cell cycle): Click or tap here to enter text.

* 1. List antibiotic resistance genes contained on these vectors: Click or tap here to enter text.
  2. If the host is a human or animal pathogen, are any of the antibiotics listed above used to treat the disease in humans or animals:  Yes  No

If yes, are any of the antibiotics used to treat the disease in any patient population? Yes No

If yes, are any of the antibiotics used to treat the disease in other countries?  Yes No

If yes, list antibiotics that can be used to treat an infection with this microorganism, taking into consideration the inserted and naturally occurring drug resistance genes: Click or tap here to enter text.

* 1. Do experiments involve large scale (> 10 L in one container) culture?  Yes No
  2. Do experiments involve cloning biological toxin gene?

If yes, name of toxin: Click or tap here to enter text. Do experiments involve transfer of an antibiotic resistance gene into the host in addition to those contained in the vectors?  Yes No

If yes, list the name of antibiotic resistance gene: Click or tap here to enter text.

# Recombinant or synthetic nucleic acids (rsNA) in human or animal tissue cell culture

# *CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Describe types of tissue cell culture used in experiments:  cell lines primary cell  tissues
  2. List all tissue cell types to be used and indicate which are continuous (c), primary (p), or explant (e) cultures:
  3. Source (species) of tissue cell cultures: Click or tap here to enter text.
  4. Indicate from where each cell line, primary cells, and/or tissue is obtained (e.g. specify name of supplier company, biobank, collaborator or own harvest protocol): Click or tap here to enter text.
  5. Gene(s) encoded by inserted DNA: Click or tap here to enter text.
  6. Biological origin of inserted DNA: Click or tap here to enter text.
  7. List Vectors to be used: Click or tap here to enter text.
  8. List antibiotics used during tissue cell culture? Click or tap here to enter text.
  9. Does vector contain > 2/3 of eukaryotic viral genome?  Yes  No

If yes, name of virus: Click or tap here to enter text.

* 1. Recombinant virus constructed?  Yes  No

If yes, Name of virus: Click or tap here to enter text.

If yes, Name of transgene: Click or tap here to enter text.

* 1. Reassortant virus constructed i.e. genetic exchange between segment viruses?  Yes  No

If yes, describe reassortant virus constructed in Overview section.

* 1. Packaging cell line used?  Yes  No

If yes, packaging cell line name: Click or tap here to enter text.

If yes, virus produced is: Click or tap here to enter text.

* 1. Defective virus with helper virus?  Yes  No

If yes, helper virus: Click or tap here to enter text.

* 1. Will inserted gene(s) be expressed?  Yes  No

If yes, what are the gene product effects (toxicity, physiological activity, oncogenic potential, or ability to alter cell cycle): Click or tap here to enter text.

# Use of lentiviral vectors

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Provide reference for vectors used (commercial source reference or published source reference) Click or tap here to enter text.
  2. Is the vector HIV-1 based?  Yes  No
  3. What is the transgene? Click or tap here to enter text.
  4. Does the transgene have oncogenic potential?  Yes  No
  5. How many plasmids are used in the packaging system? Click or tap here to enter text.
  6. Has expression of TAT been deleted?  Yes  No
  7. Is the HIV ENV gene present in the packaging system?  Yes  No
  8. If the HIV envelope gene has been replaced, does the new envelope protein broaden the host cell and tissue tropism of the lentivirus?  Yes  No
  9. If animals are involved, is lentivirus injected into the animal?  Yes  No
  10. Is the animal engrafted with transduced cells?  Yes  No

# Generation of transgenic animals (including knockouts) by embryo injection

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Species of animal: Click or tap here to enter text.
  2. Gene or sequence injected: Click or tap here to enter text.
  3. Biological origin of inserted DNA: Click or tap here to enter text.
  4. What are the anticipated effects of inserted DNA (e.g. toxicity, physiological activity, oncogenic potential, ability to alter cell cycle): Click or tap here to enter text.
  5. Describe the precautions that will be taken to minimize the possibility that animals could escape confinement: Click or tap here to enter text.
  6. Describe the effect on the wild population if an animal were to escape and mate with wild-type:

Click or tap here to enter text.

# Generation of transgenic animals by breeding

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Species of animal: Click or tap here to enter text.
  2. Are two different transgenic/knockout animal strains bred to create a new strain?  Yes  No
  3. Is a transgenic (or knockout) animal strain being bred to a non-transgenic strain?  Yes  No
  4. Does either transgenic parent contain a transgene under the control of a gamma-retroviral long terminal repeat (LTR)?  Yes  No
  5. Does either transgenic parent contain more than 50% of the genome of an exogenous eukaryotic virus from a single family?  Yes  No
  6. Is the transgenic animal that results from this breeding expected to contain more than one-half of exogenous viral genome from a single family of viruses?  Yes  No

1. Describe the expected unique characteristics of viable offspring: Click or tap here to enter text.
2. Describe the precautions that will be taken to minimize the possibility that animals could escape confinement: Click or tap here to enter text.
3. Describe the effect on the wild population if an animal were to escape and mate with wild-type:

Click or tap here to enter text.

# Recombinant or synthetic nucleic acids (rsNA) in animals

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Animals as host for recombinant microorganism?  Yes  No

If yes, name of recombinant microorganism: Click or tap here to enter text.

If yes, name of vector: Click or tap here to enter text.

If yes, name of inserted gene: Click or tap here to enter text.

If yes, biological origin of inserted gene: Click or tap here to enter text.

* 1. Animals as host for rsNA (e.g. modified plasmids)?  Yes  No

If yes, name of vector: Click or tap here to enter text.

If yes, name of inserted gene: Click or tap here to enter text.

If yes, biological origin of inserted gene: Click or tap here to enter text.

* 1. Animals as host for recombinant eukaryotic cells?  Yes  No

If yes, species of cells: Click or tap here to enter text.

If yes, name of vector: Click or tap here to enter text.

If yes, name of inserted gene: Click or tap here to enter text.

If yes, biological origin of inserted gene: Click or tap here to enter text.

# Recombinant or synthetic nucleic acids (rsNA) in insects

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Construction of transgenic insects?  Yes  No

If yes, Insect species: Click or tap here to enter text.

If yes, Transgene: Click or tap here to enter text.

If yes, biological origin of transgene: Click or tap here to enter text.

If yes, Vector: Click or tap here to enter text.

* 1. Insects as host for rsNA-modified microorganisms?  Yes  No

If yes, Insect species: Click or tap here to enter text.

If yes, Recombinant microorganism: Click or tap here to enter text.

If yes, Gene(s) encoded by inserted DNA: Click or tap here to enter text.

If yes, biological origin of inserted DNA: Click or tap here to enter text.

If yes, Vector: Click or tap here to enter text.

# Recombinant or synthetic nucleic acids (rsNA) in plants

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Construction of transgenic plants?  Yes  No

If yes, Plant species: Click or tap here to enter text.

If yes, Transgene: Click or tap here to enter text.

If yes, Biological origin of transgene: Click or tap here to enter text.

If yes, Vector: Click or tap here to enter text.

* 1. Plants as host for rsNA-modified microorganisms?  Yes  No

If yes, Plant species: Click or tap here to enter text.

If yes, Recombinant microorganism: Click or tap here to enter text.

If yes, Gene(s) encoded by inserted DNA: Click or tap here to enter text.

If yes, Biological origin of inserted DNA: Click or tap here to enter text.

* 1. Where will infected plants be located? Greenhouse Field Other: Click or tap here to enter text.
  2. Describe procedures for containment of infected plants: Click or tap here to enter text.
  3. Describe how infected plant materials rendered non-infectious: Click or tap here to enter text.

# Gene editing technologies (e.g., CRISPR/Cas9, TALEN, Zinc Finger Nucleases, Meganucleases, or any other gene editing technology)

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Please list the technology used: Click or tap here to enter text.
  2. If CRISPR is involved, are the guide RNA sequence and the Cas endonuclease on the same plasmid or delivery vehicle? Yes (**Explain in Overview**) No

If yes, can the plasmid, vector or delivery vehicle infect a human cell? Yes  No

* 1. Does the project involve a viral vector? Yes (**Explain in Overview**) No
  2. Is this a gene drive experiment? Yes (**Explain in Overview**) No
  3. Will the research involve embryos or germ line cells? (outside of standard transgenic animal protocols?) Yes (**Explain in Overview**) No
  4. Discuss the potential effects off-target events (changes in viability, morphology, physiology?): Click or tap here to enter text.
  5. How many genes have been targeted?

Single

Multiple? Specifically, how many? Click or tap here to enter text.

Library? List numbers, i.e. hundreds, thousands, more? Click or tap here to enter text.

Number of unique vectors associated with gene editing library: Click or tap here to enter text.

Number of gene editing sequences targeting each gene in the library (per vector): Click or tap here to enter text.

# Toxins

*CHECK IF THIS SECTION IS NOT APPLICABLE*

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* 1. Name of Toxin: Click or tap here to enter text.
  2. Subtype(s): Click or tap here to enter text.
  3. Active toxin? Yes (**Explain in Overview**) No
  4. Name of organism (Genus species) that produces the toxin: Click or tap here to enter text.
  5. LD50 in humans or rodents: Click or tap here to enter text.
  6. Form purchased (e.g., solid/liquid): Click or tap here to enter text.
  7. Maximum quantity in possession at any one time: Click or tap here to enter text.
  8. Is toxin a Select Agent?  Yes  No
  9. If yes, is maximum quantity below allowable limit?  Yes  No
  10. How is toxin inactivated? : heat chemical radiation

Other: Click or tap here to enter text.

# Biohazardous agent

*CHECK IF THIS SECTION IS NOT APPLICABLE*

*CHECK IF THIS SECTION IS NOT APPLICABLE*

1. Name of organism: Click or tap here to enter text.
2. Strain, subtype, serotype: Click or tap here to enter text.
3. Is antibiotic resistance expressed?  Yes  No
4. Name of antibiotics to which strain is resistant: Click or tap here to enter text.
5. Name of antibiotics to which strain is sensitive: Click or tap here to enter text.
6. Is toxin produced?  Yes  No

If yes, Name of toxin(s) produce: Click or tap here to enter text.

If yes, is work conducted with the toxin? Click or tap here to enter text.

1. Largest volume of organism cultured: Click or tap here to enter text.
2. Is organism concentrated?  Yes  No
3. If yes, method used to concentrate organism: Click or tap here to enter text.
4. How is the organism inactivated?  Heat Chemical  Radiation

Other: Click or tap here to enter text.

# Human blood, human cell culture, or unfixed human tissue

*CHECK IF THIS SECTION IS NOT APPLICABLE*

*CHECK IF THIS SECTION IS NOT APPLICABLE*

1. Describe Samples manipulated: Blood Serum Unfixed Tissue Cell culture

Other: Click or tap here to enter text..

1. List all human cells and/or tissues to be used and indicate which are continuous (c), primary (p), or explant (e) cultures: Click or tap here to enter text.
2. Indicate from where each human cell line, primary cells, and/or tissue is obtained (e.g. specify name of supplier company, biobank, collaborator or own harvest protocol) Click or tap here to enter text.
3. If human cell culture used list the source of these cells: Click or tap here to enter text..
4. Are procedures done in a Biological Safety Cabinet (BCS)? Yes No
5. If yes, provide specific BSC information in *SECTION III* Question 1
6. Are sharps used?(e.g. needles, scalpels, Pasteur pipets, razor blades) Yes No
7. Are sharps disposed into Biohazardous sharps containers? Yes No
8. Have all personnel completed annually required bloodborne pathogens training? Yes No Please provide documentation of BBP training as an Appendix to this document.
9. Are human subjects involved in this research? Yes No
10. If yes, has this work been registered with the Institutional Review Board (IRB)? Yes No
11. If no, why not? Click or tap here to enter text.
12. If yes, provide applicable **IRB Protocol(s) numbers**. Additionally, include any determination protocol numbers for non-human subjects made by the IRB, clearly indicating each type. Click or tap here to enter text.

# Non-human primate (NHP) blood, or unfixed tissue

*CHECK IF THIS SECTION IS NOT APPLICABLE*

*CHECK IF THIS SECTION IS NOT APPLICABLE*

* 1. Samples manipulated: Blood Serum Unfixed Tissue Cell culture.

Other: Click or tap here to enter text.

* 1. Are procedures done in a biological safety cabinet? Yes No
  2. Are sharps used? (e.g., needles, scalpels, Pasteur pipettes) Yes No

Are sharps disposed into red Biohazardous sharps containers Yes No

* 1. Have all personnel who work with NHP material been educated on the risks of

*Cercopithecine herpesvirus I* (monkey B virus)? Yes No

e. Are animal subjects work involved in this research? Yes No

f. If yes, has this work been registered with the Institutional Animal Care & Use Committee (IACUC)?

Yes No

If no, why not? Click or tap here to enter text.

If yes, provide applicable **IACUC Protocol(s) numbers** Click or tap here to enter text.

# Animal blood, cell culture, or unfixed tissue

*CHECK IF THIS SECTION IS NOT APPLICABLE*

*CHECK IF THIS SECTION IS NOT APPLICABLE*

1. Describe Samples manipulated: Blood Serum Unfixed Tissue Cell culture

Other: Click or tap here to enter text..

1. List all animal cells and/or tissues to be used and indicate which are continuous (c), primary (p)

or explant (e) cultures: Click or tap here to enter text.

1. Indicate from where each animal cell line, primary cells, and/or tissue is obtained (e.g. specify name of supplier company, biobank, collaborator or own harvest protocol) Click or tap here to enter text.
2. Are procedures done in a Biological Safety Cabinet (BCS)? Yes No
3. If yes, provide specific BSC information in *SECTION III* Question 1
4. Are sharps used?(e.g. needles, scalpels, Pasteur pipets, razor blades) Yes No
5. Are sharps disposed into Biohazardous sharps containers? Yes No
6. Have all personnel who work with animal material been educated on the risks of animals harboring a variety of viral, bacterial, parasitic, and fungal agents. Yes No
7. Have all personnel completed annually required bloodborne pathogens training? Yes No Please provide documentation of BBP training as an Appendix to this document.
8. Are animal subjects work involved in this research? Yes No
9. If yes, has this work been registered with the Institutional Animal Care & Use Committee (IACUC)? Yes No
10. If no, why not? Click or tap here to enter text.
11. If yes, provide applicable IACUC Protocol(s) numbers Click or tap here to enter text.

*Please complete Section III, regardless of your selections in Section II.*

## SECTION III

1. **Containment equipment available:**

Biological safety cabinet: Yes  No

Location of BSC used (Building and Room): Click or tap here to enter text.

BSC ID number: Click or tap here to enter text.

Date of last BSC certification: Click or tap here to enter text.

Containment centrifuge used :  Yes No

Location of containment centrifuge: Click or tap here to enter text.

Other containment equipment used: Click or tap here to enter text.

List Shared/Core Facility if used: Click or tap here to enter text.

1. **If live vertebrate animals are involved:**

Animal species: Click or tap here to enter text.

Animal housing location(s): Click or tap here to enter text.

1. **Overview.** Please describe the experimental design and purpose/objectives of your research, highlighting the methodology used and/or the use of infectious microorganisms if applicable.

Please write in layman’s terms but **be specific**. *Do not cut and paste from a grant application.*

Click or tap here to enter text.

1. **Risk Assessment and Control.** Please provide a protocol-specific risk assessment.

Include consideration of agent pathogenicity, virulence, infectious dose, route of transmission, host range, and stability, as well as the likelihood of exposure and consequences of exposure.

How will identified risks be controlled (e.g., engineering controls, work practices, PPE etc.)?

Include how potentially infectious wastes will be collected and disposed.

Are there specific processes that should not be conducted by a lone worker? i.e. buddy system required for safety.

Click or tap here to enter text.

1. **Training.** Please describe ALL training that will be provided to lab staff (EHS provided, online, lab specific, core facility required). This should include potential for zoonoses, or lab acquired infections, considerations for staff with immunocompromising conditions, proper operation of lab equipment, general safety information, reporting requirements, etc. Is there a process for working under supervision until competency is achieved?

How is training documented?

Click or tap here to enter text.

1. **Post-exposure procedures.** Please describe procedures that will be followed in the event of an

accidental release or exposure, including first aid, spill cleanup and reporting requirements to supervisor, EHS incident reporting if no injury, Workplace Injury Reporting to SRS Occupational Health and Safety if staff injured.

**Note:** Spills or accidents resulting in release or potential exposure to rsNA must be reported to the UO Biosafety Officer in compliance with the requirements of the *NIH Guidelines*.

Click or tap here to enter text.

**7. Proposed Containment and Practices: BSL1 BSL2 BSL1 Plant**

**ABSL1 ABSL2 BSL2 Plant**

**8. Please select all applicable section(s) of the *NIH Guidelines*:**

|  |  |
| --- | --- |
| ***Sections III-A,B,C: Experiments requiring NIH and IBC approval PRIOR to initiation:*** | |
|  | III-A: Deliberate transfer of a drug resistance trait to microorganisms that are not known to acquire the trait naturally if such acquisition could compromise the use of the drug to control disease. |
|  | III-B: Cloning of toxin molecules with LD50 of less than 100 ng / kilogram body weight. |
|  | III-C: Transfer of rsNA to human research participants. |
| ***Section III-D - Experiments that require IBC approval PRIOR to initiation****:* | |
|  | III-D-1: Experiments using [Risk Group 2, 3, or 4](http://osp.od.nih.gov/sites/default/files/NIH_Guidelines.html#_Toc351276293) agents as host-vector systems. |
|  | III-D-2: Experiments in which DNA from [Risk Group 2, 3, or 4](http://osp.od.nih.gov/sites/default/files/NIH_Guidelines.html#_Toc351276293) agents is cloned into nonpathogenic  prokaryotic or lower eukaryotic host-vector systems. |
|  | III-D-3: Experiments involving the use of recombinant or reassortant viruses in tissue culture systems; or defective recombinant viruses in the presence of helper virus or packaging cells in tissue culture systems  (this includes all eukaryotic viruses). |
|  | III-D-4-a: Experiments that generate transgenic animals, including insects (with the exception of transgenic rodents requiring BL1 containment. See III-E-3 and III-F). |
|  | III-D-4-b: Experiments involving viable rsNA-modified microorganisms tested on whole animals. |
|  | III-D-5: Experiments involving whole plants that require BL3 or BL4 containment. |
|  | III-D-6: Experiments involving more than 10 liters of culture. |
|  | III-D-7: Experiments involving human influenza strains H2N2, 1918 H1N1, and/or highly pathogenic H5N1. |
| ***Section III-E - Experiments that require registration simultaneous with initiation:*** | |
|  | III-E-1: Introduction into cultured cells of any rsNA containing greater than half but less than 2/3 of a eukaryotic viral genome (with the exception of Risk Group 3 or 4 agents). |
|  | III-E-2: Experiments involving whole plants that require BL1 or BL2 containment. |
|  | III-E-3: Generation of rodents in which the animal's genome has been altered by rsNA, requiring BL1 containment. |
|  | III-E: Experiments not specified on this sheet. |
| ***Section III-F - Experiments that are exempt but still require registration:*** | |
|  | III-F-1: Experiments that use synthetic nucleic acids that can neither replicate nor generate nucleic acids capable of replicating in any living cell; are not designed to integrate into DNA, and do not produce a toxin that is lethal for vertebrates at an LD50 of <100 ng/kg body weight. |
|  | III-F-8: Cloning of all other DNA in *E. coli* K12, *S. cerevisiae*, and *B. subtilis* host-vector systems (with the exception of DNA from Risk Group 3 or 4 pathogens). |
|  | III-F-8: Introduction into cultured cells of any rsNA containing less than half of a eukaryotic viral genome (with the exception of Risk Group 3 or 4 pathogens). |
|  | III-F-8: Purchase or transfer of transgenic rodents that require BL1. (Appendix C-VII) |
|  | III-F-8: Breeding experiments to generate transgenic rodents that may be housed under BL1, with the exception of those listed in Section III-E. (Appendix C-VIII) |

***Principal Investigator:*** *“By submitting this registration, I accept responsibility for the safe conduct of work with this material at the established biosafety level. I will ensure that all personnel receive training in regard to proper safety practices and personal protective equipment needed for this work.”*

***Signed\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_***

Version updated April 2025